Exhibit 1

9/21 Sequencing gel of SEC-2, T7, 1259C 3398

Transfection of DC plasmids

		μl used
Water		10
pcDNA7		10
pcDNA7 FT3m		10
pcDNA7 FT3m, F6,3	(3+2)	4
pcDNA7 FT3m, F6,4	(5+9)	12
pcDNA7 FT3m F6L	(I-1)	5
pcDNA7 FT6m F3,4	(12+3)	20
pcDNA7 FT6m F3,345		

Standard DEAE Dextran transfection protocol on COS-7 cells.

DNA harvested for use in above Transfection

	260	280	C	260/280
5+9	.255	.137	2.5	1.86
12+3	.237	.143	2.4	1.65
12-7	.244	.144	2.4	1.70

9/22 long gel of SEC-2 samples
SEP5 9925
SEP6 1259, T7, 3348B

Begin Sequencing FT7 Clone – 104 Sal/EcoRI

Primers

8993	3244 (NOT WORK)
714	8904
946	8902
7931	8875
8661A	8851
8953	8771

9/23 Sequencing gel (Formamide Gel 40%) of the above Samples

11/1 TA cloning (Invitrogen)   Followed instructions to letter

	P/F	P+	insert
10X	1.0	1.0	
Vec	2.0	2.0	
Lig	1.0	—	
H_2O	5.0	6.0	
	1.0	1.0	10pg
E			

11/2 Transformation

lig	1.0	1.0	1.0 ctrl
Cells	50	50	50
Broth	450	450	450

thaw cells
2µl β MEOH
Mix by TAP
Add DNA

Also transform
KG's pcR products cloned into pcDNA1
- MC[illegible]61/P3

Set up PCR experiment (in KG Project)
use Vector primers ; 3rd exon primers
to evaluate 14-7 FD DNA from library (PLS) pcD
2nd Amp 1 µg/µl
this will tell if insert is in the library

Primers
5315 } pcDM7 Vector primers
5316 }
152
153

11/1 Completed the assembly and checking of any sequence discrepancies in FT7 Sequence
This is 3594 nt Sequence
from 300 → end in 2 directions
Gave this sequence to S.N. to check against his independently determined sequence
11/2 no differences detected - full 99.9% confident this sequence